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(54) **Regulated genes by stimulation of chondrocytes with IL-1beta**

(57) The present invention refers to the novel use of osteopontin, calnexin and TSG-6 gene product in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues and to novel genes induced or repressed by stimulation of chondrocytes with IL-1 β and their use in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues.

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Description

The present invention refers to the novel use of osteopontin and calnexin in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues and to novel genes induced or repressed by stimulation of chondrocytes with IL-1 β and their use in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues.

Among the diverse biological effect of interleukin-1 (IL-1), are its actions on the metabolism of many connective tissue cell types including articular chondrocytes. IL-1 inhibits proteoglycan (PG) synthesis by chondrocytes and stimulates production of prostaglandin E₂ and metallo-proteinases capable of degrading matrix macromolecules. From experimental results, and from findings of IL-1, PG fragments and proteolytic enzymes in inflamed joints, it was deduced that IL-1 plays a role in cartilage degradation in osteoarthritis and rheumatoid arthritis (Benton HP & Tyler JA. 1988, Biochem. Biophys. Res. Comm. 154, 421-428; Aydelotte MB et al. Conn. Tiss. Res. 28, 143-159; Wood DD et al., Arthritis Rheum. 26, 975-983; Lohmander LS et al., Trans Orthop. Res. Soc. 17, 273). Matrix metalloproteinases are potential candidates for drug interaction at the enzyme level, but relevant molecular targets interfering with earlier processes leading to cartilage degradation are still lacking. Therefore, one objective of the present invention was to identify potential targets for drug modification of IL-1 β induced cartilage degradation on the RNA level of human articular chondrocytes from osteoarthritic cartilage.

As an initial attempt to investigate differentially expressed genes in diseased cartilage, total RNA from IL-1 β stimulated and unstimulated human chondrocytes was subjected to differential display of mRNA by reverse transcription and polymerase chain reaction (DDRT-PCR). This method can be used to identify and isolate those genes that are differentially expressed in two cell populations (Liang P & Pardee AB 1992, Science 257, 967-971; Liang P et al., AB 1993, Nucl. Acids Res. 21, 3269-3275; Bauer D et al. 1993, Nucl. Acids Res. 21, 4272-4280). The key element is to use a set of oligonucleotide primers, one hybridizing to the polyadenylated tail of mRNAs, the other being arbitrary decamers that anneal at different positions relative to the first primer. mRNA subpopulations defined by these primer pairs are amplified after reverse transcription and resolved on DNA sequencing gels. Band patterns are created, which are characteristic for each RNA population extracted from the cell population under study. For example, 100 different primer combinations should generate a total of approximately 10,000 PCR products for each population, which should represent about the half of all expressed cellular genes. A comparison of the band pattern obtained from two cell populations reveals differentially displayed bands which correspond to differentially expressed genes. Subsequently, differentially displayed bands can be extracted from the gel, reamplified, subcloned and sequenced.

Due to its extreme sensitivity, the appearance of artifactual bands is an inherent problem of the DDRT-PCR method used according to the present application. An additional problem is also the evaluation of complex gene expression patterns. Yet another problem of the present invention is that only minute amounts of RNA are available.

Therefore, it was particularly surprising that the DNA TAU1/1 with the sequences

TAU1/1(1)

ACATCACCTC	ACACATGGAA	AGCGAGGAGT	TGAATGGTGC	ATACAAGGCC	ATCCCCGTTT	60
CCCAGGACCT	GAACCCGCCT	TCTGATTGGG	ACAGCCGTGG	GAAGGACAGT	TATGAAACGA	120
GTCAGCTGGA	TGACCAGAGT	GCTGAAACCC	ACAGCCACAA	GCAGTCCAGA	TTATATAAGC	180
GGAAA						185

and

TAU1/1(2)

CTAAATGCAA	AGTGAGAAAT	TGTATTTTTT	CTCCTTTTAA	TTGACCTCAG	AAGATGCACT	60
ATCTAATTCA	TGAGAAATAC	GAAATTCAG	GTGTTTATCT	TCTTCCTTAC	TTTTGGGG	118

and the DNA TTU2/2 with the sequence

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5  AACCAGTATT TCAAACTAT TATCTGGATT CAAGATTAGT GTGTAAAGAT TGTTTTCTTA 60
    TCAGTAAAT AGGTCTTCAG ATCTGCATCT GGCCTCTTAG CATGTTTTTC TTCATAGATA 120
    CCCGTTTTGG GGTTTTTGCG TCGGAAGATG AATGGCATT ATAGTCCTCT CCACATTAT 180
    CTG 183

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are 100 % identical to human osteopontin cDNA and 97.2 identical to human calnexin, respectively. This demonstrates that the experimental approach of the present invention worked efficiently, i.e. the use of 100 different primer combinations (25 oligodecamer primers, 4T₁₂MN-primers) generated a total of approximately 10.000 PCR products for each population which represent 53 % of all expressed cellular genes. 123 PCR bands out of 10.000 appeared as differentially expressed bands. 53 of the original 123 PCR bands were reproducibly displayed by comparing the PCR band patterns from two patients; of those 68 % arose from IL-1 β stimulated chondrocytes.

It was further found that osteopontin which is a secreted highly acidic phosphoprotein of 32 kd (Denhardt and Guo (1993) FASEB J. 7, 1475-1482) is surprisingly downregulated in IL-1 β stimulated human chondrocytes. This means that osteopontin is involved in IL-1 β related diseases of connective tissues, in particular osteoarthritis.

Osteoarthritis is characterized as a slowly progressing matrix degeneration with continuing degradation of collagens and proteoglycans and subsequent release of matrix fragments into the synovial fluid. Any disturbance of the normal chondrocyte matrix interactions, for example through a loss of osteopontin, could cause an altered signaling through the integrin $\alpha_5\beta_1$ and thus changed cellular responses leading to early steps of matrix degradation.

Therefore, one embodiment of the present invention is the use of osteopontin itself or parts thereof, antibodies against it or nucleic acids such as DNA or RNA or parts thereof coding for osteopontin or parts thereof in the diagnoses, prophylaxis or therapy of IL-1 β related diseases of connective tissues, in particular osteoarthritis. According to the present application the term "parts" means either at least 8, preferably 12, in particular 15 amino acids in case of proteins or 6-100, preferably 10-40, in particular 12-25 nucleic acids in case of DNA or RNA as hybridization probes. The methods of diagnosing such diseases will be described infra. In addition, quantification on the protein level is possible with osteopontin specific antibodies on Western blots, in immunocytochemistry, FACS analysis or ELISA based assay systems. The present invention refers also to a diagnosis aid or a pharmaceutical for such use. Osteopontin can be produced for example recombinantly through expression in procaryotes, in insect cells in mammalian cells or in mammalian cells using Vaccinia as detailed in Ausubel et al. 1994 [Current protocols in molecular biology, Chapter 16, John Wiley & Sons, Inc]. The cDNA of Osteopontin is e.g. disclosed in Young et al. (1990), Genomics 7, 491 - 502.

Antibodies against osteopontin can be generally produced for example by the method of Neil GA & Urnovitz HB (Trends in Biotechnology, 6, 209-213, 1988) or Köhler G & Milstein C (Nature, 256, 52-58, 1975).

Also calnexin which is an integral membrane protein of 88 kd (Bergeron et al. (1994) TIBS 19, 124-128) is surprisingly downregulated in IL-1 β stimulated human chondrocytes compared to unstimulated chondrocytes. This means also that calnexin is involved in IL-1 β related diseases of connective tissues, in particular osteoarthritis. In addition, a downregulation of the calnexin synthesis would cause a reduced amount of correctly and completely folded proteoglycans because calnexin is a new type of molecular chaperone that associates with incompletely folded proteins such as proteoglycans. Proteoglycans are highly glycosylated glycoproteins which are of central importance for the maintenance of the cartilage tissue integrity.

Hence, an additional embodiment of the present invention is the use of calnexin itself, or parts thereof antibodies against it or nucleic acids such as DNA or RNA or fragments thereof coding for calnexin or parts thereof in the diagnosis, prophylaxis or therapy of IL-1 β related diseases of connective tissues, in particular osteoarthritis. The methods of diagnosing such diseases are already described above. The present invention refers also to a diagnosis aid or a pharmaceutical for such use.

Calnexin can be produced for example recombinantly as described above for osteopontin. The cDNA of Calnexin is e.g. disclosed in Galvin et al. (1992), Proc. Natl. Acad. Sci. USA 89, 8452 - 8456. The production of said antibodies are also generally described above.

Potential role of identified cDNA fragments in IL-1 mediated cellular processes TSG-6

A homology search in the GenBank and EMBL databases revealed a 99.5 % sequence identity of fragment TAU7/2(c) with the gene coding for human TSG-6. TSG-6 (TNF stimulated gene 6) was originally isolated by differential cDNA library screening as a TNF induced gene sequence from human fibroblasts (Lee et al., 1990). It was further characterized by Lee et al (1992) as a TNF and IL-1 inducible, secretory, 39 kDa glycoprotein with extensive sequence homology with a region implicated in hyaluronate binding, present in cartilage link protein, proteoglycan core proteins,

and the adhesion receptor CD44. With the ability to bind HA and with the most extensive sequence homology to CD44, TSG-6 belongs to the hyaladherin family. Wisniewski et al. (1993) detected high levels of TSG-6 protein in synovial fluids of patients with various forms of arthritis. Six normal control patients did not contain detectable TSG-6 protein in their joint fluid, whereas joint fluids from nine rheumatoid arthritis patients contained high, moderate or low levels of TSG-6. Two patients with osteoarthritis had high levels of TSG-6 in their joint fluids. The apparent local source of TSG-6 in the joints are synoviocytes and chondrocytes (Wisniewski et al., 1993). Lee et al. (1992) speculated that TSG-6 could act as a competitive inhibitor of the interaction between CD44 and its ligand(s) and thus might influence the structural organization of the extracellular matrix of connective tissue, resulting in a destabilization of the proteoglycan aggregates.

Hence, an additional embodiment of the present invention is the use of TSG-6 gene product itself, or parts thereof antibodies against it or nucleic acids such as DNA or RNA or fragments thereof coding for TSG-6 gene product or parts thereof in the diagnosis, prophylaxis or therapy of IL-1 β related diseases of connective tissues, in particular osteoarthritis. The methods of diagnosing such diseases are already described above. The present invention refers also to a diagnosis aid or a pharmaceutical for such use.

Fibronectin

A homology search in the GenBank and EMBL databases revealed a 100 % sequence identity of fragment TTO20/1(c) with the gene coding for human fibronectin.

Fibronectin is a 450 kd glycoprotein with various functions. It acts as an adhesive ligand, as growth or differentiation factor and has chemotactic properties. It is found in the extracellular matrix of most types of cells (Hynes R 1993. Fibronectins, In: Guidebook to the extracellular matrix and adhesion proteins. Editors: Kreis T, Vale R. Oxford University Press. 56-58). An enhanced accumulation of fibronectin and fragments derived from it are found in the synovial fluid and on the inflamed synovial and pannus surfaces in the knee joint of patients with rheumatoid arthritis (Dutu A, Vlaicu-Rus V, Bolosiu HD, Parasca I, Cristea A. 1986. Fibronectin in plasma and synovial fluid of patients with rheumatic diseases. Med. Interne 24, 61-68). Patients with osteoarthritis, as well, have greatly increased levels of fibronectin in their synovial fluid and on cartilage surfaces (Xie D-L, Meyers R, Homandberg GA. 1992. Fibronectin fragments in osteoarthritic synovial fluid. J. Rheumatology 19, 1448-1452). The intraarticular injection of fibronectin fragments causes a severe depletion of cartilage proteoglycans in vivo (Homandberg GA, Meyers R, Williams JM. 1993. Intraarticular injection of fibronectin fragments causes severe depletion of cartilage proteoglycans in vivo. J. of Rheumatology 20, 1378-1382), which is explained by the induced release of several proteinases, including stromelysin (Xie D-L, Hui F, Meyers R, Homandberg GA. 1994. Cartilage chondrolysis by fibronectin fragments is associated with release of several proteinases: Stromelysin plays a major role in chondrolysis. Arch. Biochem. and Biophysics 311, 205-212). At high concentrations, fibronectin fragments enhance cartilage catabolism through release of cytokines, including IL-1 (Homandberg et al., personal communication).

In respect to these published data, the upregulation of fibronectin by IL-1 can be regarded as a positive feedback regulation, enhancing the self destructive potential of chondrocytes and synoviocytes. With this, fibronectin expression is a direct pharmacological target.

In addition, the sequencing of differentially displayed PCR products discovered also unknown DNA fragments which correspond to differentially expressed genes with or without stimulation of chondrocytes with IL-1 β .

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Therefore, another embodiment of the present invention is a DNA containing a DNA selected from the group consisting of

5 TA08/2(2)
 1 CCAAGTTTT CCAGCAACCC CAAGGGAATA CAGGGAGATC AATGCACCA
 51 AAATGGGAAA AGAAAAATAC TTCGATGCAA TGAAACAAAG CCTTTTCCG
 101 TTCAGTTTCC ATAATTTCAGT GGTCAGTTT AAGGCTGCCA CTTGGG

10 TA016/1(2)
 1 GACACGAACA CCACATATTT TTATTGGAGG CCCCATGGCT CCTTGGGAAGC
 51 CATTTTGGAA CCAAGGGGAC CCACCTTTTT

15 TA016/2(2)
 1 CTAAATATAT TCTCTAACAA GTTAATCTCT TTCAAATCTA TAGATAAAAC
 51 TAAAGGATA AGGAACCAAG GTTTAACCGA CCTAGCCAAT TATGGCAATC
 20 101 ATACTTGCTT TTTAG

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TA017(C)

5 1 CATGAAATAT TTCTTGAGGT AATAAGCTTT TACCAAGCTT ATATTTTGG
51 GCAATTCAGT TACAATGAGA AAAAAACACA CCAAAGACC AAAAATTTTA

101 AAAACTCACT TTTCTTGCAA TCATAGACAT TTGCATTATT ATAGAACATT
151 CAAACAAGTT AGGTGGATAA TTATTGTCTA TAGATAAATA CGATGCAATT
110 201 TTTTAAATGT ATGACCGATA CTCCGTATAT ACTTAGATAA CTTATCCAGA
301 AACCTCAACT GTATTGAACA TTGCTGAGAG AAATCAACAA TAATTTTAAC
351 AGATATGATG ACAGNAAAAA TTGATTGCAT ATCTTTTGGC ACTAAACTT
401 TTATATTTAT TT

15 TA019(C)

1 AGAGCAGGGG TATTTNCNGG TTCATACCGC CATGGCTTAA GAAGCAAAAG
51 TCATATACCT TAGTAGTGGC AAAGATNGAG GAGATAAAAA AGAGCCTACC
20 101 CAAGCTGTTG TTGAAGAACA GGTCTTAGAT AAAGAGGAAC CCTTCCAGAA
151 GNACAGAGAC AGGCTAAGGG TGATGCTGAG GAAATGGCTC AGAAGAAACA
201 AGAGATTAA

25 TAU 1/1(2)

1 CTAAATGCAA AGTGAGAAAT TGTATTTTT CTCCTTTTAA TTGACCTCAG
51 AAGATGCACT ATCTAATTCA TGAGAAATAC GAAATTCAG GTGTTTATCT
101 TCTTCCTTAC TTTTGGGG

30 TAU 1/1(1)

1 ACATCACCTC ACACATGGAA AGCGAGGAGT TGAATGGTGC ATACAAGGCC
51 ATCCCCGTTT CCCAGGACCT GAACCCGCCT TCTGATTGGG ACAGCCGTGG
101 GAAGGACACT TATGAAACGA GTCAGCTGGA TGACCAGAGT GCTGAAACCC
35 151 ACAGCCACAA GCAGTCCAGA TTATATAAGC GGAA

TAU1/2(2)

1 CCGGAATGGG GAGCAAACCTA TAAGAACCGG GACCAGTTTC CTCTCTTTGT
51 GCCCTAGTTC CCCCTCCTTT GTATACACCC TCCATCCTGA ATAGACTCTG
40 101 GTTCTCAGCG TAACACCGAC AACATTCAAT CCTGTAGAGA AACAAATGTT
151 AGCTCAGAAG GACACAGCCT TTGAATCATC AGAGAGTT

TAU 7/1(2)

45 1 GTTAAGAATA ACTAAATAAA AGTTTTAATT AATTTAGGAA TATAAAAAAC
51 TATTAACATT TAATTTTATA ACTGTATCTG CCAAGCAACT TTAAATATAA
101 TTTATTTACC

50 TAU 7/1(1)

1 CACGCAATGT GAAATAGGCA CATAGGAAGA ATGGGGAAAC CATCCCCTCA
51 AGCATTTATC CTTTGAGTTA CAAGCAATCC AATTACACTC TTTTAGTTAT
101 TTTTAAATGT ACAGTTAGGT TATTA

55

TAU 7/2(C)

5
1 CCTTGAAGAT GACCCAGGTT NCTTGGCTGA TTATGTTGAA ATATAGACA
51 GTTACGATGA TGTCCATGGC TTTGTGGGAA GATACTGTGG AGATGAGCTT
101 CCAGATGACA TCATCAGTAC AGGAAATGTC ATGACCTTGA AGTTTCTAAG
151 TGATGCTTCA GTGACAGCTG GAGGTTTCCA AATCAAATAT GTTGCAATGG
201 AT

TAU10(1)

1 GGAGATGACA TTTGCTTTGG GCAGAGGCAG CTAGCCAGGA CACATTTCCA
51 CTATAATTTT ACAAAGTTAA ATTTATAAGC TAGCATTAAAG TAAAGTGAAG
15 101 TTCCAGCTCC CTGCTAAAA ATAAC TAGAG GTAATAATTG GTATT CAGGT
151 AACTCATTTA CATCATAATG TGTGTGAAA A

TAU12/1(2)

20 1 TATAAAATAT AAATTATATT ATAAATCATG TATTATTTAT AAAATTATAT
51 TATAAATTTA TAAAAATATA AATTATATTT TAGGCTTAAT GTATAAGGAA
101 TATAAATTAT TAATAAGCAT ATGA

TAU 12/1(1)

26 1 TGTAATTAAC TGINCTTGTA GGTCTGTCTT TTATACATGT GTGAGTTTTT
51 CTTTACAATA GATTCCTAGC ATTGGGATG CTAGGTCAGA TGGTATGCAC
101 ATTTGACATT TTGATTGATA GCACCAGATT GCTTTGTTAA AAAATTTTNN
30 151 TTTATAGTTT ACATTATCTT TGTACAATAG ATGTTCTCTT TCGAC

TAU 12/2(1)

1 GGGAAGTGAA TTGAAAATAC TTCTTTNTCA ACATAATTTT NGGGTTTTGA
51 AATTGTGTTT GGGTTTTTCAG GAAATTGGTG GTAATCTTGT ATTAGACTGAA
35 101 AAAAAGTGAA TTTTAAATTT CTCAGTGAAG AAGCAAATGA TTTATTTTTC
151 ATAGA

TAU12/3(2)

40 1 TGTTCGTGTA ACTGTTCTAA TTGTGTCTTT GTTACTTCCA GTGCAACCCCT
51 TTCAGGTAAG

TAU12/3(1)

45 1 CTAAAGAACT TGGTATCTCT ATTAAAGCAC ACGAACCTCC AAGGAAAATA
51 GAGCGATTTA CTCTTCTCAT ATCAGTGCAT ATTTATAAGA AGCACGGAGT
101 CA

TAU13/1(1)

50 1 AGTCATCAAT TCCTTTTTAT CTGTAATTAC ACATTGTTT TTATTTCAAA
51 GTAATTATAA GGTGTTATAT TGCATATAAT CAGAAAACTA AATGGAAATA
101 AAATTTTAGT AAGCCCGGCC CCTTTGACCG ATACAGAAAA CTGA

55

TAU 13/3(2)

5 1 TATATGGCAG TCTAAAGCAT CAAAGATTG CATCAACATC TTTCATTTTA
 51 GACATCTCCT TGCAATGTAA AATATCATGT ATCAACAACA TCTGGTGCAA
 101 ATCCATGAGT CTAACTCGAC ATTTCATCTA GCTCGATTAT TATTCCTTCG
 151 TACAGTCGAT GTAAACAATA CAGAAAGAGG ATTATTAAGA ACAGTTT

TAU 13/3(1)

10 1 ATTCATGAAA TGGTCTATAT GCATGATATT GTAAATTCGG ACTCGAAACC
 51 GAAACCAAGG ATTCCGTTAC AAAAATTCCT TAATGCTGAG AATGTTCTCA
 101 CGCAAACAAC ATCATGGACA TTAAATTCAA GATATGTGAA TGTTAATTCT
 151 GTCAATAAAG TCAACGTAAA GAGTAAAGTT AAAAACAGTT ATATCTNNNC
 15 201 TGTCATGAT GAGTTTAGTT TAACAGATGA TGAATCAATT CT

TCO 16/1(C)

20 1 CAAAGTGT TTGGTTTGA GAGAGAGAGA GATTGAGAGA CAGAGAGAGA
 51 GAGAGAAACC AAGGGATCAT GATAGTTATA GTCAAATACG AGGTTGGATT
 101 ATCTTTTGAA AATGTGTGG TTCTGTGATA CAAGAGGAAG CTAAGACATA
 151 TCGTGGAAC ATCTCCCCC TCCACCTTAA TATCAAGAAC AAATTGTGGA
 201 ATCTAATGTT AATGAGAAGT AGTTCCCCAC TGTGTCAGAT G

TCO16/2(C)

25 1 NCATCTGACA CAGTGGGGAA CTACTTCTCA TTAACATTAG ATTCCACAAT
 51 TTNNNCTTGA TATTAAGGNN NNNNNGGGAG ATCGTTTCAC GATATCGTCT
 101 TAGCTTCCTC TTGTATCACA GAACCAACAC ATTTCAAAG ATAATCCTTC
 30 151 CTCNNTTGA CTATAACTAT CATGATCCCT TGGTCTCTC TCTCTCTCTG
 201 CTCTCTCATC TCTCTCTCTC TNAAAACNAA

TCO17(C)

35 1 ACAGTAGTTA GGAGTTTCTT TACTTACAAA ATCACTGGAA ATGATTAAAT
 51 TGCTTTTCCC CCTCCCCAGA GGTGCATTTT TCTTATTTC ATATAGTAAA
 101 GTTGAGCTTT TACAGTGCAT AATGTGACAT TTGGAATGCT TATCAACTGC
 151 ATGTAAACAT TAATAACCT

TCO18(C)

40 1 GTAAATGGTA TTANNNGCTG AAGAAAAAAA ATTTTTCAG ACCTCTGTTT
 51 TTTAACTGAA CTTTATCATT GGCATTGTGG GCTTTGAAGT TGCTGGGATA
 45 101 AATTAATATA ATTAATAAAA AGACTGAATT TAATTGCAAA AAAAAAAAAA
 151 AACAATAAGT GTGGTGAT

TCU2/1(1)

50 1 AAGAAATTAT CCAGTTATTT ACAAGGCCAC TGATATTTTA AACGTCCAAA
 51 AGTTTGTTTA AATGGGCTGT TACCGCTGAG AATGATGAGG ATGACAATGA
 101 TGTTGAAGG TTACATTTTA GGAAATGAAG AACTTAGAA AATTAATATA
 151 AAGACAGTGA TAAATACAAA GAAGATTT

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TCU2/2(1)

5 1 CGGGTTAATA TTATCCTCTA GTATAAGTGA ATTACTAGTT TCTCTTTATT
51 TAGACAAACA CACACACACC AGATAATATA AACTTAATAA ATTATCTGTT
101 AATGTAGATT TTATTTAAAA AACTATATTT GAACATTGGT CTTTCTTGGA
151 C

TCU9/1(2)

10 1 ACATAACAGC TTTTATACAA TGATAAGGAC ATATCATTTG TTTACAAAGA
51 AAGTCTAAAA TTTCAGAAGC ATTCAAAGAG CTAACACAGT AAAGGTCATG
101 CAAGTTCTAG AATAGTGAAT CATGACAGAA CTCATTCAAT TTATCCTTTA
151 TCTCC

TCU9/2(2)

20 1 AAGTATGGGT AGCTAAATTT GCATTAAATT AAAAGTACAT ATAATGCAAC
51 ACCACTCTAC ATCTGTATAC CTACGAATGT ATGTGTACTA CACACCCCTTA
101 AAATGTTTTT CAAAGTCTTA ATATATTAGA ACATGTTTTT ATTTTTTCAT
151 GGGATGTTAA TACTATTCTA TGATTAAGAA AATACTAG

TCU10(2)

25 1 AATACAGTTA TTCTAGCTTT TCATATTCAA TTTGAATGAT CAGAAAAGTA
51 TATTAGTCAC ACAGAATTAA ATATTTTACA TAGTAAGAAT C

TCU14(2)

30 1 GAAGTGAAAG TCAGCCCTTT AGCTATTATT TATTGCTTTA TTAGAGCAGA
51 GGGAAAGTGAC ACTCATTGCC TTCACAGAGC TCTGCAGAAA TATATGCACA
101 GAGTGGTCAA TGCCAACATC TGAGTAAGTC TTCCAAA

TGO20(2)

35 1 CAGAACATTA GGATTTATTC CTTGATTAGT TCAAATGATT TCAACAGCTG
51 AATTCCTTGA GATGTGTAAG GCAGGTTGGT CCTTTGGATG GACTGTAGAC
101 TGAAACTTCC TATAACTGTA GTGATATGTA CACAGCTACA TAGCAAAGTG
40 151 CTTCAATTATG AAAATGAAGA A

TGO20(1)

45 1 CAGTGTGAGA GTCTCATTTT TATGCACAGT GTTTCTCAGG AGGATGGAGC
51 TAGTTAGCTG TCTGTTGTCT GTAGCCAGC TTGATAATGG AACTATACAG
101 CGAAGAGACA ATCTCTGGCA AGTTTTTGTG GAA

TGU5(C)

50 1 TTAGAGTAAA ATTCCAAATA AATGCTTTGC TCCAAAATTA CACTAACCAG
51 GCTGGGTCTC TATCATACAT CTTCAATACC CTCAAACCTA GATTGTAAAG
101 TGAAAAAAGT GATTAGCNNT TCCATTTGTT CATTCGTGCA CTCACATTCT
151 TAGGCATTTT AAGGATGAGC AACCTTTGTT TCAGAAAGGG TAAGTAATTA
201 GCCCCCTGGA GGTACATAG TTATAATTTA GTCTTCAGAA TCCGTTTCGAA

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251 GGGNNNNGTT ACTATTTTTA AGATAATTAG AACCCACCTT GTAGCAATAA
 301 AAGTTTTCTT GTCTTTG

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TGU8(2)

1 GCGAAAGACT AATCGAACCA TCTAGTAGCT GGTTCCTCC GAAGTTTCCC
 51 TCAGGATA

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TGU9/1(2)

1 TTAATGTTTA AATACTACTT TTTTTCAG CTGCCCCTAG ATACCAACTG
 51 TTTATCTAAC ACACAATTCC AGTGTGCGCA AGCCTCATGC CAATTTGAAG
 101 GGAACAGCCA AAACCTTATGC ATTCATATAA AAAGAGTCTC TAGGCTCTTA
 151 TATCTACATT ATAATTTTT

15

TGU9/2(2)

1 GGAATAACAT TTTTATGA GGAACCCCTT TAAATGGAT GCACACAGTG
 51 GCATTTTCTC CTAGGCTCAA AGCTGAGTAC ACTCCCGTAA TTTTAATAAT
 101 ATTTTAGGCA AGTCCTATGA CAATTATACC AACAGTTTC TTCAACCCCA
 151 CCACCACCCC ACCATCTCTA TGC

20

TGU12(C)

1 GGAGGAAGCT TTATTGGGA AGAGTGGGT TCNNTCGGCC CTGATCAGCT
 51 CTAGCCTGCC CACCCCATCT CAGCCAGGCG GCTTTACTTC TTCTGAGCT
 101 TCAGGTCTTT CTTCTTCTG ATTTCTTGG CCAGCTCCCC AATCAATCTC
 151 CAGTACTCAT TGAACCTGAG CTCGAGNCC TGATTCACAT CCAAGCTCTT
 201 CATCTTCT

25

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TGU13/1(C)

1 GGATGTGGTA GTTGATCTTT AATGCCCAT CTAGGTGGGA AAAATCCATG
 51 ATCCTAATT TTAAGAGAAG GTTGGTAACT CTACTTAGGA CTTTTTTTG
 101 TAAGAGGAAT AATGTAGCCT CACCCCTATC TTTCTGGAAA TGTTTAAACC
 151 ACTGAAATAT GGAGATCAA TCCAGCTTAC ACACTGGTAA CTCAAATACT
 201 ATTTTTTTTT TAACTATCT TTTCTAACT AATCACCCT CTTGTACATA
 251 GAACCTTCTA TCTCAGTGCC AATTCTTAGA GGTGATGCA AACAGCTCTC
 301 CAGAGAGCCT GTGCTATTGT TC

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TGU13/2(2)

1 GGGGTGTACA TTTTATTGGA AACCTTAAAT ACTGTTGAGA AAGAATATAT
 51 CTTCAATCAA GGTCTTGCCG AGCCTACACA GAAAAATGAA GCTTTTGGG
 101 TTAGGGGCAA GGATATATAC AGTACAGAGG ACAAAGA

45

TT016/2(C)

1 ACATTCATTA AAGATGAACT TTCAGCATCT TCACTTGAAG ATCCATCAGA
 51 TGATTCTGAG AGGCAGGTCT CCCCCAAAAA TCCACCGCAT GTATTCTTTC
 101 GTTTAGAATC TGAAAGCCTC TTTCTTTTCA GGCTTGATGA CTCTTCTAAG
 151 GTATTTGTTA TGCCTCTCTT CTGGGTTTTT CGTTTGCCT TATCAAGTAG

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	201	CTNAAATTCA	AACACCATGG	CAANAGAAAC	TGCTTCTAT	
5		TTO20/1(2)				
	1	CCACCAGCCT	ACTGATCAGC	TGGGATGCTC	CTGCTGTCAC	AGTGAGATAT
	51	TACAGGATCA	CTTACGGAGA	AACAGGAGGA	AATAGCCCTG	TCCAGGAGTT
	101	CACTGTGCCT	GGGAGCAAGT	CTACAGCTAC	CATCAGCGGC	CTTAAACCTG
10	151	GAGTTGATTA	TACCATCACT	GTGTATGCTG	TCACTGGCCG	TGGAGACAGC
	201	CCCGCAAGCA	GCAAGCCAAT	TTCCATTAAT	TACCGAACAG	AAATTGACAA
	251	ACCATCCCAG	ATGCAAGTGA	CCGATGTTCA	AGACAACTGT	TTTAATAAAA
	301	GATTTACATT	CCAC			
15		TTO20/2(2)				
	1	TTGGTACCAC	AGTCACAGAA	CTGGGGGTCA	TTTTCTAGAT	GAAACAAACG
	51	GAACAAGTTC	TCTTCCAACA	AAGAAATGTA	CTGTAGAAAT	TAATTTCCCTC
20	101	CATGAATTTT	ATATATTGTG	TACAAATATA	AGGTATGTAT	CTGAATACAA
	151	AG				
		TTU2/1(2)				
25	1	CTAGAACTTC	CAAAGGCTGC	TTGTCATAGA	AGCCATTGCA	TCTATAAAGC
	51	AACGGCTCCT	GTTAAATGGT	ATCTCCTTTC	TGAGGCTCCT	ACTAAAAGTC
	101	ATTTGTTACC	TAAACCTTAT	GTGCCTTAAC	AGGCCAATGC	TTCTCG
		TTU 2/2(C)				
30	1	AACCACTATT	TCAAACTAT	TATCTGGATT	CAAGATTAGT	GTGTAAGAT
	51	TGTTTTCTTA	TCAGTAAAAT	AGGTCTTCAG	ATCTGCATCT	GGCCTCTTAG
	101	CATGTTTTTC	TTCATAGATA	CCCGTTTTGG	GGTTTTTGCG	TCGGAAGATG
	151	AAGTGCAGTT	TATAGTCCTC	TCCACATTTA	TCTG	
35		TTU3(1)				
	1	GGGTAGAAAG	CTGAATAATT	TATGAAGGAG	AGGGGTCAGG	GTTGATTCGG
	51	GAGGACCTAT	TGCTCGGGGG	GCTTTGTATG	ATTATGGGCG	TTGATTAGTA
40	101	GTAGTTACTG	GTTGAACATT	GTTTGTGGT	GTATATATTG	TAATTGAGAT
	151	TGCTCGGGGG	AATAGGTTAT	GTGATTAGGA	GTAGGGTTAG	GATGAGTGGG
	201	AAG				
		TTU 5/1(2)				
45	1	GACAAAAAAA	AAAAAACAGG	TTTAAAGCT	AGAAATGAAA	AGCTACTTAA
	51	GTATCTTAAA	GGATAAGTTA	CTTTATTATA	CACTAGAAAC	ATACACAATA
	101	GCTGAAAAC	TAAAAATCT	CACACTGCTG	AATGTCTCTG	CTGGCTG
		TTU5/2(2)				
50	1	GCATCCATTG	TACATTGTTT	GGTTTGAGGT	TACCATGAGG	CCTGTAAATA
	51	CTATCTTATA	ATTTATTATT	TCAACCTGAT	AAAACCTAAC	ACTATTTGCA
	101	TAAACAAACA	AACGAAAA			
55						

TTU9/1 (1)

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1 TAAATACTG GTTCTTTTAT TCTGCAATAT TTTTAAAAAT CACATTTTCA
51 GCCAGGCGCA GTTTCCACACA CCTGTAATCC GGCACTTTGG GAGGCTGAGA
101 TGGGTGGATC ACAAGGTAGG AGATCAAACA TCCTGGCCAA CATGGTGAAC
151 CTGTTTACT

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TTU9/2 (2)

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1 CAAGTATGGG TAGCTAAATT TGCATTAAAT TAAAAGTACA TATAATGCAA
51 CACCACTCTA CATCTGTATA CCTACGAATG TATGTGTACT ACACACCCTT
101 AAATGTTTCA AAGCTTAATA TATTAGAACA TGTTTTTCATT TTCAGGGAG

TTU13 (2)

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1 GGAAATACAC TAGCATGTGA GCACTGTATA TAAAGCTTGA GGTTAGGAGG
51 TAAATGAAA GAAATCATTT TTAACCTCTA AGATGT

TTU13 (1)

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1 TGAATTAAAT GGACTCGTTG AAAGGACAAG GAGATCGGTA ATATCTCTCT
51 AAAGAACTTA TATACTAAAA TCTGTAATTG CCTGTACCAA AAGTTTTAGT
101 CTTCTTTT

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or an analog thereof. In accordance with the invention, the term "analog" includes nucleic acids which code for the same protein sequence due to the degeneration of the genetic code, for a protein having conservative amino acids substitutions or deletions that do not eliminate the characteristic feature of this protein, or for a protein having at least about 85 %, and more advantageously at least about 90 %, in particular 95 % amino acid sequence homology.

Other embodiments of the invention provide a vector containing said DNA and a host cell containing said vector.

According to the general knowledge one skilled in the art can also use said nucleic acids of the present invention as a hybridization probe to detect the corresponding genes in an organism or in a sample from an organism or gene mutations thereof.

Therefore, an additional embodiment is a method for isolating a gene which can be induced or repressed by treating chondrocytes that contain this gene by IL-1 β containing the steps:

45 (a) hybridizing a DNA of the present invention under stringent preferably high stringent conditions against DNA or RNA containing said gene, preferably DNA or RNA isolated originally from chondrocytes, in particular human chondrocytes; and

(b) isolating this gene by methods known to a skilled person in the art.

50 According to the present invention the term "stringent conditions" means hybridization conditions comprising a salt concentration of 4 x SSC (NaCl-citrate buffer) at 62-66°C, and "high stringent conditions" means hybridization conditions comprising a salt concentration of 0,1 x SSC at 68°C. The length of the probes are 6-100, preferably 10-40, in particular 12-25 nucleic acids long.

Yet another embodiment is a process for expressing a gene isolated according to the above-described process containing the steps:

55 (a) cloning said gene into a suitable expression vector such as the pET series (Studier et al., 1990. Methods in Enzymology 185, 60) for procaryotic expression or the vector CDM8 for mammalian expression (Aruffo and Seed, 1987. Proc. Natl. Acad. Sci. USA 84, 8573) or any other expression system known to one skilled in the art; and

(b) expressing said gene in a suitable host cell such as BL21 series (Studier et al., 1990, supra) for procaryotic expression or COS, cells for mammalian expression (Aruffo and Seed, 1987, supra) or any other expression system known to one skilled in the art;

5 or a method for producing a protein containing the steps:

(a) culturing a suitable host cell, in particular the above mentioned, containing a vector, in particular an expression vector such as the vectors mentioned above which contains a DNA or a gene of the present invention; and

10 (b) isolating the expressed protein for example by ultrafiltration, precipitation with chaotropic agents such as urea or column chromatography on e.g. ion exchange chromatography columns as detailed in Ausubel et al. 1994 (supra).

A further embodiment is a diagnostic aid containing a DNA or parts thereof or a gene or parts thereof of the present invention. In particular, quantification of the genes can be achieved on the RNA level by Northern blotting with gene specific probes of the present invention or with gene specific primers in a PCR reaction. Such primers can be synthetically produced using the DNA sequences of the present invention or the sequences of the corresponding genes. Therefore, said nucleic acids are useful for the diagnosis of IL-1 β related diseases of connective tissues, in particular osteoarthritis or rheumatoid arthritis.

These nucleic acids can also be used to evaluate the expression of certain genes in small cartilage biopsies and to use these ultimately as disease-specific markers and/or as predictive markers for disease progression of e.g. osteoarthritis. The hybridization conditions can be the same as described above.

Said nucleic acids, however, can also be used for the therapy against the diseases mentioned or for the production of a pharmaceutical.

Therefore, another embodiment of the present invention is also the use of said nucleic acids for the production of a pharmaceutical. For example, as described by Uhlmann & Peyman (Chem. Rev. (1990), 90, 543), Milligan et al. (J. Med. Chem. (1993), 36, 1923) or Stein & Cheng (Science (1993), 261, 1004) such nucleic acids can be used as antisense oligonucleotides or triple helix forming oligonucleotides for the inhibition of gene expression. This is in particular useful if such a disease is caused by the overproduction of a gene product which is directly or indirectly regulated by IL-1 β in chondrocytes. The nucleic acids can additionally be modified in order to increase e.g. the stability against nucleases as described e.g. in the literatures mentioned above.

Finally, also the gene product itself produced by a method of the present invention can be used as a pharmaceutical.

In the following the invention is in particular described by the examples and tables:

Description of the Tables

Table 1 gives an overview on used primers and the complexity of the detected differences in expression.

Table 2 summarizes the result of the sequencing of differentially displayed PCR products after their elution from the sequencing gel, reamplification and subcloning into the pCRII vector. The sequences of TAU1/1(1) and TAU1/1(2) are 100 % identical to human osteopontin cDNA, the sequence of TTU2/2 is 97.2 % identical to human calnexin. bp = base pairs, IL-1 = Interleukin-1 stimulation, Stat. sig. score = statistical significance score: a feature of the BLAST database searching program. This score is determined using an implementation of Karlin's significance formula (Karlin, S. and Altschul, S.F. 1990. Methods for assessing the statistical significance of molecular sequence features by using general scoring schemes. Proc. Natl. Acad. Sci. USA, 87:2264-2268), which calculates the Poisson probability that the observed sequence similarity will occur by chance based on the size and composition of the sequence database as well as on the size and quality of the match. The smaller this number, the more it is likely to see sequence similarities.

Examples

Cell culture

Articular cartilage specimen were obtained from two patients (male 65 years old and female 73 years old) undergoing total joint replacement surgery for osteoarthritis. None of these individuals had received treatment by radiation or chemotherapy. Articular cartilage slices were aseptically dissected from both femoral condyles, tibia plateaus and patellae and subjected to sequential enzymatic digestion with pronase and collagenase as described (Häuselmann HJ et al. 1992, Matrix 12, 116-129) Since it is known that the alginate gel suspension system retains the chondrogenic phenotype [Lohmander LS et al. 1992, Trans. Orthop. Res. Soc. 17, 273.] 4×10^6 chondrocytes were suspended in low viscosity alginate (4×10^6 cells / ml 1,25 % w/v alginate in an isotonic buffered solution) and expressed through a 22 gauge needle into 102 mM CaCl solution to form cell entrapping beads which are 1,5-3 mm in diameter and spherical. Alginate beads containing a total number of 2×10^7 cells were fed daily for the first three days with medium F12 / DMEM (50/50)

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and 10 % fetal calf serum (Sigma) with 25 µg / ml ascorbate and 50 µg / ml gentamycin and were then subdivided into two populations for further three culture days in the presence or absence of 5U / ml rh IL-1β (Genzyme). For cell recovery, alginate beads were finally dissolved into dissolution buffer (55 mM sodiumcitrate, 30 mM EDTA, 0,15 M NaCl) and placed at room temperature for 10 min. Viability was checked by eosin-red exclusion and cell number was determined.

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Primer syntheses

Arbitrary oligodecamer primers OPA6 to OPA10, OPA16 to OPA20 and degenerate anchored oligo-dT primers (T₁₂VN) were synthesized using the 392 DNA synthesizer (Applied Biosystems) and purified by denaturing polyacrylamid gel electrophoresis. Some oligodecamer primers, U1 to U15 were purchased from Biometra (Göttingen, FRG).

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List of all degenerate 3' oligo dT-primers [T₁₂VN] used for DDRT-PCR:

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Primer	Sequence 5' to 3'
T ₁₂ VA	5'-TTTTTTTTTTTTTVA-3'
T ₁₂ VA	5'-TTTTTTTTTTTTT VT-3'
T ₁₂ VA	5'-TATTTTTTTTTTTVG-3'
T ₁₂ VA	5'-TTTTTTTTTTTTTVC-3'
V = dA, dG, dC; N = dA, dT, dG, dC	

List of all arbitrary 5' oligodecamer primers used for DDRT-PCR:

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Primer	Sequence 5' to 3'
OPA 6	GGTCCCTGAC
OPA 7	GAAACGGGTG
OPA 8	GTGACGGGTG
OPA 9	GCGTAACGCC
OPA 10	GTGATCGCAG
OPA 16	AGCCAGCGAA
OPA 17	GACCGCTTGT
OPA 18	AGGTGACCGT
OPA 19	CAAACGTCCG
OPA 20	GTTGCGATCC
U1	TACAACGAGG
U2	TGGATTGGTC
U3	CTTTCTACCC
U4	TTTTGGCTCC
U5	GGAACCAATC
U6	AAACTCCGTC
U7	TCGATACAGG
U8	TGGTAAAGGG
U9	TCGGTCATAG
U10	GGTACTAAGG
U11	TACCTAAGCG
U12	CTGCTTGATG
U13	GTTTTCGCAG
U14	GATCAAGTCC
U15	GATCCAGTAC

RNA isolation and cDNA synthesis

Total RNA from cultured articular chondrocytes was prepared according to a single step method Chomczynski and Sacchi (Chomczynski P & Sacchi N 1987, Anal. Biochem. 162, 156-159) and incubated with 10 U RNasefree DNaseI (Gibco, Eggenstein, FRG) for 30 min at 37°C to remove chromosomal DNA contamination of RNA. After extraction with phenol/chloroform (3:1), the supernatant was ethanol precipitated in the presence of 0.3 M NaOAc and RNA was redissolved in DEPC treated water. 0.4 µg total RNA was then reverse transcribed using 200 U M-MLV (Moloney murine leukemia virus) reverse transcriptase (Gibco, Eggenstein, FRG) in a 40 µl reaction volume containing 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 10 mM DTT, 3 mM MgCl₂, dNTP mix (dATP, dTTP, dCTP, dGTP) of 200 µM each, 40 U RNase inhibitor (Boehringer Mannheim, FRG) and 2.5 mM degenerate oligo-dT primer (T₁₂VN) at 37°C for 1 h. Reactions were terminated by heating for 5 min at 95°C.

PCR amplification

cDNAs were amplified in a DNA thermal cycler (Perkin Elmer, model 480) in 20 µl PCR reactions containing 2.5 µM of the original T₁₂MN-primer used in cDNA synthesis in combination with 0.5 µM arbitrary upstream primer, dNTP mix (dGTP, dCTP, dTTP) of 0.5 µM each, 10 µCi α-[³⁵S]dATP (1000 Ci/mmol, 10 mCi/ml), 10 mM Tris-HCl (pH 8.3) 50 mM KCl, 1.5 mM MgCl₂, 0.001 % gelatin and 2.5 U AmpliTaq DNA polymerase. Light mineral oil was overlaid and thermal cycling was performed as follows: 94°C for 30 seconds, 40°C for 2 min and 72°C for 30 seconds for 40 cycles followed by 5 min postextension at 72°C. AmpliTaq DNA polymerase was purchased from Perkin-Elmer (Weiterstadt, FRG) and α-[³⁵S]dATP was obtained from Amersham-Buchler (Braunschweig, FRG). After addition of 5 µl stop buffer (95 % formamide, 20 mM EDTA, 0.05 % bromophenolblue and 0.05 % xylene cyanol) radiolabeled PCR-fragments were then displayed on 6 % acrylamide/7 M Urea high resolution sequencing gels of 35 x 43 cm in size; dried gels were exposed to X-ray film (Kodak X-OMAT) and exposed for 48 h, which allows rapid identification of differentially expressed genes by side by side comparison of DDRT-PCR band patterns.

Elution, reamplification and cloning of PCR fragments

PCR fragments identified as differentially expressed bands were cut from acrylamide gels, transferred into Eppendorf tubes and rehydrated for 10 min with 100 µl 10 mM Tris-HCl and 1 mM EDTA at room temperature. After boiling the gel slice for 15 min, the PCR fragment was recovered by ethanol precipitation in the presence of 0.3 M NaAc and 20 µg glycogen as a carrier and redissolved in 10 µl sterile water. 5 µl of this volume was used for reamplification by PCR using appropriate primers and conditions described above except for dNTP concentration of 20 µM and no radioisotope. The reamplified PCR product was visualized by electrophoresis on a 2 % agarose gel and eluted from the gel by ultrafiltration using Ultrafree MC-filters (Millipore). Purified PCR fragments were then cloned into the pCR11-vector (Invitrogen, De Schelp, NL) by the TA cloning method (Kovalic D et al. 1991, Nucleic Acids Research 19, 4640), which allows in-vitro transcription and sequencing from the plasmid.

Sequencing

Plasmid DNA sequencing of subcloned PCR fragments with either SP6(2) or T7(1) primer was carried out using the chain-termination DNA sequencing method, as described by Sanger et al. (Sanger F et al. 1977, Proc. Natl. Acad. Sci. USA 74, 5463-5467.).

Sequence analysis

The sequence analysis revealed the sequences of cDNA clones TAO8/2(2), TAO16/1(2), TAO16/2(2), TAO17(c), TAO19(c), TAU1/1(2), TAU1/1(1), TAU1/2(2), TAU7/1(2), TAU7/1(1), TAU7/2(c), TAU10(1), TAU12/1(2), TAU12/1(1), TAU12/2(1), TAU12/3(2), TAU12/3(1), TAU13/1(1), TAU13/3(2), TAU13/3(1), TCO16/1(c), TCO16/2(c), TCO17(c), TCO18(c), TCU2/1(1), TCU2/2(1), TCU9/1(2), TCU9/2(2), TCU10(2), TCU14(1), TCU14(2), TGO20(2), TGO20(1), TGU5(c), TGU8(2), TGU9/1(2), TGU9/2(2), TGU12(c), TGU13/1(c), TGU13/2(2), TTO16/2(c), TTO20/1(c), TTO20/2(2), TTU2/1(2), TTU2/2(c), TTU3(1), TTU5/1(2), TTU5/2(2), TTU9/1(1), TTU9/2(2), TTU13(2), TTU13(1) disclosed on pages 7 to 14 of the specification.

Searching for homology between subcloned PCR fragments and sequences already listed in one of the DNA databases (GenBank or EMBL database) was performed using the FASTA program developed by Pearson and Lipman (Pearson W & Lipman DJ 1988, Proc. Natl. Acad. Sci. USA 85, 2444-2448) included in the GCG software package (Genetics Computer Group, Madison, USA).

Northern-blot analysis

Cell culture and isolation of RNA was performed exactly as described above. 10 µg of total RNA from both IL-1β stimulated or not stimulated chondrocytes were denatured by heating at 65°C for 10 min in a solution of 50 % formamide, 20 mM MOPS and 2.2 M formaldehyde, separated through a 1 % agarose gel containing 2.2 M formaldehyde in 1 X MOPS and transferred to positively charged nylon membrane (Amersham) by standard blotting procedures [Maniatis et. al 1992]. After UV crosslinking, the blots were prehybridized for 1 h in rapid-hyb-buffer (Amersham) at 65°C. A 330 bp cDNA corresponding to nts 61 to 390 of human osteopontin cDNA (GenBank J04765) and a 340 bp cDNA corresponding to nts 881 to 1220 from human calnexin (GenBank M94859) were radiolabeled for hybridization with α-[³²P]dCTP (3000 Ci/mmol, 10 mCi/ml) using random nonamer primers (Amersham) up to a specific activity of ~ 1,5 x 10⁹ dpm / µg DNA. Hybridization was performed for 2,5 h at 65°C in prehybridization solution with 2 ng / ml of labeled probe added. The blot was subsequently washed in 2 X SSC, 0.1 % SDS at 37°C for 15 min (1 X SSC = 0,15 M NaCl, 0.015 M sodium citrate, pH 7,0), followed by two successive washes with 1 X SSC, 0.1 % SDS at 65°C for 10 min respectively. If necessary, a final high stringency wash was performed with 0.1 X SSC, 0.1 % SDS at 65°C for 15 min. The blots were then analysed by autoradiography using Kodak X-Omat films at -80°C with intensifying screens for 2-7 days and intensity of bands was quantified with a phosphorimager (Biorad, model GS-250). All blots were stripped with boiling 0.5 % SDS solution and re probed with labeled β-actin to demonstrate equal loading of RNA in each lane.

Northern hybridisations (Results)

Fragment TAU7/2(c), identical to TSG-6, was differentially upregulated in IL-1 stimulated cells. This is in concordance with Lee et al. (1992) which reported for TSG-6 a TNF-α and IL-1 mediated upregulation. Fragment TAU1/1, identical to human osteopontin and fragment TTU2/2, identical to human calnexin, both were weaker expressed in IL-1 stimulated chondrocytes compared with the unstimulated cells. To validate our differential display data, we performed Northern analyses of Osteopontin and calnexin expression in IL-1 stimulated and unstimulated chondrocytes originating from a third patient. Both messages were again downregulated. A phosphorimager quantification revealed an osteopontin downregulation by 79% and a calnexin downregulation by 40% in the RNA population from chondrocytes of the third

Table 1: Current results of differential display reverse transcriptase PCR (DDRT-PCR) to reveal differential gene expression by chondrocytes with and without IL-1 β

Overview on used primers and number of analysed bands

DDRT-PCR primercombination

3'-Oligo dT-primer (downstreamprimer)	5'-Oligodecamer (upstreamprimer)	putative differential expressed genes by side by side comparison DDRT-PCR band pattern	reproducibility of DDRT-PCR band pattern from first to second, third or fourth DDRT-PCR (same patient ¹) (other patient ²)		eluted from gel and reamplified in PCR	cloned into pCRII vector by TA cloning method verified by PCR	PCR-fragment sequenced using SP6 or T7 promoter
T ₁₂ M*A	OPA 6 - OPA 10	25 out of ~ 4000 bands	7	not done	8	1	1
T ₁₂ M*T	OPA 16 - OPA 20	19 out of ~ 4000 bands	13	9	12	12	12
T ₁₂ M*G	U 1 - U 5	31 out of ~ 4000 bands	not done	11	11	10	10
T ₁₂ M*C	U 6 - U 10	27 out of ~ 4000 bands	not done	13	12	11	11
	U 11 - U 15	21 out of ~ 4000 bands	not done	11	11	10	10
total 4 x = 100 combinations	25	total 123	total	55	total 52	total 44	total 44

* means threefold degeneracy where M may be dA, dG or dC

¹ patient female 73 years old diagnosis gonarthrosis

² patient male between 65-75 years old

theoretical consideration:

Suggesting that an arbitrary upstream primer detects 3 % of the total RNAs (Liang 1994), then 97 % of the total mRNAs will not be detected, i.e. with 25 arbitrary oligodecamerprimer and the four degenerate T₁₂VN primers, about half of the mRNAs would be seen ($P = 1 - (0,97)^n = 1 - (0,97)^{25} = 53,3 \%$) in 100 lanes of high resolution sequencing gels.

Table 2 IL-1 mediated differentially displayed cDNA fragments of human articular chondrocytes

Fragment	bp	IL-1	Features	Stat.sig.score
TAO 8/2(2)	275 bp	+	146 bp sequenced, no homology found	0.999
TAO 16/1(2)	450 bp	+	80 bp sequenced, no homology found	0.69
TAO 16/2(2)	200 bp	+	115 bp sequenced, no homology found	0.04
TAO 17(c)	412 bp	+	412 bp sequenced, no homology found	0.016
TAO 19(c)	209 bp	--	209 bp sequenced, no homology found	0.99
TAU 1/1(1,2)	450 bp	--	100 % sequence identity to human osteopontin cDNA in 303 bp overlap (303 bp seq.)	1.2×10^{-101}
TAU 1/2(2)	430 bp	+	188 bp sequenced, no homology found	0.82
TAU 7/1(1,2)	500 bp	+	87 % sequence identity to human cDNA clone c-1sd02 in 125 bp overlap (235 bp seq.)	8.1×10^{-33}
TAU 7/2(c)	202 bp	+	99.5 % sequence id to human TNF stimulated gene-6 in 202 bp overlap	4.8×10^{-76}
TAU 10(1)	400 bp	+	181 bp sequenced, no homology found	0.9997
TAU 12/1(1,2)	470 bp	--	319 bp sequenced, no homology found	3.3×10^{-14}
TAU 12/2(1)	390 bp	--	155 bp sequenced, no homology found	0.0078
TAU 12/3(1,2)	250 bp	--	95 % sequence identity to human cDNA clone HRBBA21 similar to S10 in 158 bp overlap (162 bp seq.)	1.0×10^{-28}
TAU 13/1(1)	600 bp	+	145 bp sequenced, no homology found	0.12
TAU 13/3(1,2)	500 bp	--	439 bp sequenced, no homology found	0.33
TCO 16/1(c)	241 bp	+	241 bp sequenced, no homology found	2.4×10^{-7}
TCO 16/2(c)	230 bp	+	230 bp sequenced, no homology found	4.3×10^{-5}
TCO 17(c)	169 bp	+	169 bp sequenced, no homology found	0.49
TCO 18(c)	168 bp	+	168 bp sequenced, no homology found	1.3×10^{-8}
TCU 2/1(1)	400 bp	+	178 bp sequenced, no homology found	0.68
TCU 2/2(1)	210 bp	+	151 bp sequenced, no homology found	0.0074
TCU 9/1(2)	430 bp	+	99 % sequence identity to human cDNA clone 131036 3' in 155 bp overlap (155 bp seq.)	7.2×10^{-58}
TCU 9/2(2)	320 bp	--	188 bp sequenced, no homology found	0.22
TCU 10(2)	320 bp	--	100 % sequence identity to human cDNA clone 26518 3' in 85 bp overlap (91 bp seq.)	2.9×10^{-28}

Fragment	bp	IL-1	Features	Stat.sig.score
TCU 14(1,2)	280 bp	+	99.3 % sequence identity to human cDNA HL60 3' directed Mbol in 249 bp overlap (249 bp seq.)	3.5×10^{-51}
TGO 20(1,2)	300 bp	+	304 bp sequenced, no homology found	0.95
TGU 5(c)	317 bp	+	317 bp sequenced, no homology found	0.088
TGU 8(2)	320 bp	+	100 % sequence identity to human 28S rRNA in 58 bp overlap (58 bp seq.)	1.4×10^{-16}
TGU 9/1(2)	280 bp	+	169 bp sequenced, no homology found	0.55
TGU 9/2(2)	220 bp	--	100 % sequence identity to human cDNA clone 12A10B in 100 bp overlap (173 bp seq.)	4.0×10^{-36}
TGU 12(c)	208 bp	--	87 % sequence identity to human cDNA clone 113442 3' in 208 bp overlap	5.5×10^{-43}
TGU 13/1(c)	322 bp	+	322 bp sequenced, no homology found	6.9×10^{-13}
TGU 13/2(2)	300 bp	--	94.9 % sequence identity to human F1 ATPase β -subunit in 137 bp overlap (137 bp seq.)	2.3×10^{-43}
TTO 16/2(c)	239 bp	+	97.5 % sequence identity to human ERCC5 in 239 bp overlap (239 bp seq.)	9.3×10^{-68}
TTO 20/1(c)	314 bp	+	100 % sequence identity to human fibronectin in 314bp overlap (314 bp seq.)	1.9×10^{-121}
TTO 20/2(2)	400 bp	+	152 bp sequenced, no homology found	0.035
TTU 2/1(2)	300 bp	--	100 % sequence identity to human cDNA clone 118470 5' in 146 bp overlap (146 bp seq.)	2.1×10^{-36}
TTU 2/2(c)	184 bp	--	99 % sequence identity to human calnexin in 184 bp overlap (184 bp seq.)	2.3×10^{-64}
TTU3(1)	400 bp	+	97 % sequence identity to human NADH-DH mtDNA subunit in 203 bp overlap (203 bp seq.)	8.6×10^{-69}
TTU 5/1(2)	300 bp	--	147 bp sequenced, no homology found	0.0065
TTU 5/2(2)	270 bp	--	118 bp sequenced, no homology found	0.035

Fragment	bp	IL-1	Features	Stat.sig.score
TTU 9/1(1)	350 bp	+	94 % sequence identity to human cDNA clone 83764 3' in 159 bp overlap (159 bp seq.)	$5,9 \times 10^{-23}$
TTU 9/2(2)	320 bp	--	149 bp sequenced, no homology found	0,22
TTU 13(1,2)	350 bp	+	194 bp sequenced, no homology found	0,57

Thus, the 44 identified fragments can be subdivided as follows:

1) 2 fragments with sequence homologies to known human genes with known roles in IL-1 mediated processes:

TAU 7/2 identical with human TNF-stimulated gene-6
TTO 20/1 identical with human fibronectin

2) 6 fragments with sequence homologies to known human genes, whose function in IL-1 mediated processes can be speculated:

TAU 1/1 identical with human osteopontin
TGU 8 identical with human 28S ribosomal RNA gene
TGU 13/2 identical with human F1 ATPase β -subunit
TTO 16/2 identical with human ERCC5
TTU 2/2 identical with human calnexin
TTU 3 identical with human NADH-DH mtDNA subunit

3) 9 fragments with sequence homologies to human genes, identified in human genome sequencing projects:

TAU 7/1 identical with human cDNA clone c-1sd02
TAU 12/3 identical with human cDNA clone HRBBA21
TCU 9/1 identical with human cDNA clone 131036 3'
TCU 10 identical with human cDNA clone 26518 3'
TCU 14 identical with human cDNA clone HL60 3' directed Mbol
TGU 9/2 identical with human cDNA clone 12A10B
TGU 12 identical with human cDNA clone 113442 3'
TTU 2/1 identical with human cDNA clone 118470 5'
TTU 9/1 identical with human cDNA clone 83764 3'

4) 27 fragments without sequence homologies to known human genes The detection of TSG-6 and fibronectin, both genes known to be upregulated by IL-1, points to the importance of those other cDNA fragments in the light of IL-1 mediated processes. Those genes very likely play roles in degenerate joint diseases, including rheumatoid and osteoarthritis and with this are interesting candidates as markers for clinical studies or as drug targets for pharmacological intervention.

Claims

- Use of osteopontin itself or parts thereof, or antibodies against osteopontin or parts thereof or nucleic acids or parts thereof coding for osteopontin or parts thereof in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis.
- Diagnostic aid for the diagnosis of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing osteopontin itself or parts thereof, or antibodies against osteopontin or parts thereof or nucleic acids or parts thereof coding for osteopontin or parts thereof.

3. Pharmaceutical for the prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing osteopontin itself or parts thereof, or antibodies against osteopontin or parts thereof or nucleic acids or parts thereof coding for osteopontin or parts thereof.
- 5 4. Use of calnexin itself or parts thereof, or antibodies against calnexin or parts thereof or nucleic acids or parts thereof coding for calnexin or parts thereof in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis.
- 10 5. Diagnostic aid for the diagnosis of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing calnexin itself or parts thereof, or antibodies against calnexin or parts thereof or nucleic acids or parts thereof coding for calnexin or parts thereof.
- 15 6. Pharmaceutical for the prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing calnexin itself or parts thereof, or antibodies against calnexin or parts thereof or nucleic acids or parts thereof coding for calnexin or parts thereof.
- 20 7. Use of TSG-6 gene product itself or parts thereof, or antibodies against TSG-6 gene product or parts thereof or nucleic acids or parts thereof coding for TSG-6 gene product or parts thereof in the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis.
8. Diagnostic aid for the diagnosis of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing TSG-6 gene product itself or parts thereof, or antibodies against TSG-6 gene product or parts thereof or nucleic acids or parts thereof coding for TSG-6 gene product or parts thereof.
- 25 9. Pharmaceutical for the prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis, containing TSG-6 gene product itself or parts thereof, or antibodies against TSG-6 gene product or parts thereof or nucleic acids or parts thereof coding for TSG-6 gene product or parts thereof.

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10. DNA containing a DNA selected from the group consisting of

TA08/2(2)

5 1 CCAAGTTTT CCAGCAACCC CAAGGGAATA CAGGGAGATC AATGCACCCA
 51 AAATGGGAAA AGAAAAATAC TTCGATGCAA TGAAACAAAG CCTTTTCCG
 101 TTCAGTTTCC ATAATTCAGT GGTCACTTTT AAGGCTGCCA CTGGG

TA016/1(2)

10 1 GACACGAACA CCACATATTT TTATTGGAGG CCCCATGGCT CCTTGAAGC
 51 CATTTTGGAA CCAAGGGGAC CCACCTTTTT

TA016/2(2)

15 1 CTAAATATAT TCTCTAACAA GTTAATCTCT TTCAAATCTA TAGATAAAAC
 51 TAAAGGATA AGGAACCAAG GTTAAACCGA CCTAGCCAAT TATGGCAATC
 101 ATACTTGCTT TTTAG

TA017(C)

20 1 CATGAAATAT TTCTTGAGGT AATAAGCTTT TACCAAGCTT ATATTTTTGG
 51 GCAATTCAGT TACAATGAGA AAAAAACACA CCAAAAGACC AAAAATTTTA
25 101 AAACTCACT TTTCTTGCAA TCATAGACAT TTGCATTATT ATAGAACATT
 151 CAAACAAGTT AGGTGGATAA TTATTGTCTA TAGATAAATA CGATGCAATT
 201 TTAATAAGAA TTTGAAGAAT GACATTAAAT GCTGTCTGAA GCCTTTGTAT
 251 TTTTTAATGT ATGACCGATA CTCCGTATAT ACTTAGATAA CTTATCCAGA
30 301 AACCTCAACT GTATTGAACA TTGCTGAGAG AAATCAACAA TAATTTTAAC

351 AGATATGATG ACAGNAAAAA TTGATTGCAT ATCTTTTTCG ACTAAAACTT
 401 TTATATTTAT TT

TAO19(C)

1 AGAGCAGGGG TATTCNCGG TTCATACCGC CATGGCTTAA GAAGCRAAAG
 51 TCATATACCT TAGTAGTGGC AAAGATNGAG GAGATAAAAA AGAGCCTACC
 101 CAAGCTGTTG TTGAAGAACA GGTCTTAGAT AAAGAGGAAC CCTTCCAGAA
 151 GNACAGAGAC AGGCTAAGGG TGATGCTGAG GAAATGGCTC AGAAGAAACA
 201 AGAGATTAA

TAU 1/1(2)

1 CTAATGCAA AGTGAGAAAT TGTATTTTTT CTCCTTTTAA TTGACCTCAG
 51 AAGATGCACT ATCTAATTCA TGAGAAATAC GAAATTTTCA GTGTTTATCT
 101 TCTTCCTTAC TTTTGGGG

TAU 1/1(1)

1 ACATCACCTC ACACATGGAA AGCGAGGAGT TGAATGGTGC ATACAAGGCC
 51 ATCCCCGTTT CCCAGGACCT GAACCCGCCT TCTGATTGGG ACAGCCGTGG
 101 GAAGGACAGT TATGAACGA GTCAGCTGGA TGACCAGAGT GCTGAAACCC
 151 ACAGCCACAA GCAGTCCAGA TTATATAAGC GGAAA

TAU1/2(2)

1 CCGGAATGGG GAGCAAATA TAAGAACCGG GACCAGTTTC CTCTCTTTGT
 51 GCCCTAGTTC CCCCTCCTTT GTATACACCC TCCATCCTGA ATAGACTCTG
 101 GTTCTCAGCG TAACACCGAC AACATTCAAT CCTGTAGAGA AACAAATGTT
 151 AGCTCAGAAG GACACAGCCT TTGAATCATC AGAGAGTT

TAU 7/1(2)

1 GTTAAGAATA ACTAAATAAA AGTTTAAATT AATTTAGGAA TATAAAAAAC
 51 TATTAACATT TAATTTTATA ACTGTATCTG CCAAGCAACT TTAAATATAA
 101 TTTATTTACC

TAU 7/1(1)

1 CACGCAATGT GAAATAGGCA CATAGGAAGA ATGGGGAAAC CATCCCCCTCA
 51 AGCATTATC CTTTGAGTTA CAAGCAATCC AATTACACTC TTTTAGTTAT
 101 TTTTAAATGT ACAGTTAGGT TATTA

TAU 7/2(C)

1 CTTGAAGAT GACCCAGGTT NCTTGGCTGA TTATGTTGAA ATATATGACA
 51 GTTACGATGA TGTCCATGGC TTTGTGGGAA GATACTGTGG AGATGAGCTT
 101 CCAGATGACA TCATCAGTAC AGGAAATGTC ATGACCTTGA AGTTTCTAAG
 151 TGATGCTTCA GTGACAGCTG GAGGTTTCCA AATCAAATAT GTTGCAATGG
 201 AT

TAU10(1)

1 GGAGATGACA TTTGCTTTGG GCAGAGGCAG CTAGCCAGGA CACATTTCCT
 51 CTATAATTTT ACAAAGTTAA ATTTATAAGC TAGCATTAG TAAAGTGAAG
 101 TTCCAGCTCC CTTGCTAAAA ATAAC TAGAG GTAATAATTG GTATTTCAGGT
 151 AACTCATTTA CATCATAATG TGTGTGAAA A

TAU12/1(2)

1 TATAAAATAT AAATTATATT ATAAATCATG TATTATTTAT AAAATTATAT
 51 TATAAATTTA TAAAAATATA AATTATATTT TAGGCTTAAT GTATAAGGAA
 101 TATAAATTAT TAATAAGCAT ATGA

TAU 12/1(1)

1 TGTAAATTAAC TGTNCTTGTA GGTCTGTCTT TTATACATGT GTGAGTTTTT
 51 CTTTACAATA GATTCCTAGC ATTGGGATTG CTAGGTCAGA TGGTATGCAC
 101 ATTTGACATT TTGATTGATA GCACCAGATT GCTTTGTTAA AAAATTTTNN
 151 TTTATAGTTT ACATTATCTT TGTACAATAG ATGTTCTCTT TCGAC

TAU 12/2(1)

1 GGGAGTGAA TTGAAAATAC TTCTTTNTCA ACATAATTTT NGGGTTTTGA
 51 AATTGTGTTT GGGTTTTTCAG GAAATTGGTG GTAATCTTGT ATTAGCTGAA
 101 AAAAGTGAA TTTTAAATTT CTCAGTGAAG AAGCAAATGA TTTATTTTTC
 151 ATAGA

TAU12/3(2)

1 TGTCTGGTA ACTGTTCTAA TTGTGTCTTT GTTACTTCCA GTGCAACCCCT
 51 TTCAGGTAAG

TAU12/3(1)

1 CTAAAGAACT TGGTATCTCT ATTAAAGCAC ACGAACCTCC AAGGAAAATA
 51 GAGCGATTTA CTCTTCTCAT ATCAGTGCAT ATTTATAAGA AGCACGGAGT
 101 CA

TAU13/1(1)

1 AGTCATCAAT TCCTTTTAT CTGTAATTAC ACATTTGTTT TTATTTCAA
 51 GTAATTATAA GGTGTTATAT TGCATATAAT CAGAAACTA AATGGAAATA
 101 AAATTTTAGT AAGCCCGGCC CCTTTGACCG ATACAGAAAA CTGA

TAU 13/3(2)

1 TATATGGCAG TCTAAAGCAT CAAAGATTG CATCAACATC TTTCATTTA
 51 GACATCTCCT TGCAATGTAA AATATCATGT ATCAACAACA TCTGGTGCAA
 101 ATCCATGAGT CTAACGAC ATTCATCTTA GCTCGATTAT TATTCCTTCG
 151 TACAGTCGAT GTAAACAATA CAGAAAGAGG ATTATTAAGA ACAGTTT

TAU 13/3(1)

5
1 ATTCATGAAA TGGTCTATAT GCATGATATT GTAAATTCGG ACTCGAAACC
51 GAAACCAAGG ATTCGGTTAC AAAAATTCCT TAATGCTGAG AATGTTCTCA
101 CGCAACAAC ATCATGGACA TTAAATTCAG GATATGTGAA TGTTAATCT
151 GTCAATAAAG TCAACGTAAA GAGTAAAGTT AAAACAGTT ATATCTNNNC
201 TGTCAATGAT GAGTTTAGTT TAACAGATGA TGAATCAATT CT

TCO 16/1(C)

1 CAAAGTGTTC TTGGTTTGA GAGAGAGAGA GATTGAGAGA CAGAGAGAGA
51 GAGAGAAACC AAGGGATCAT GATAGTTATA GTCAATACG AGGTGGGATT
15
101 ATCTTTTGAA AATGTGTGG TTCTGTGATA CAAGAGGAAG CTAAGACATA
151 TCGTGGAAAC ATCTCCCCC TCCACCTTAA TATCAAGAAC AAATTGTGGA
201 ATCTAATGTT AATGAGAAGT AGTCCCCAC TGTGTCAGAT G

TCO16/2(C)

20
1 NCATCTGACA CAGTGGGAA CTACTTCTCA TTAACATTAG ATTCCACAAT
51 TNNNNCTTGA TATTAAGGNN NNNNNGGAG ATCGTTTCAC GATATCGTCT
101 TAGCTTCCTC TTGTATCACA GAACCAACAC ATTTCAAAG ATAATCCTTC
151 CTCNNTTGA CTATACTAT CATGATCCCT TGGTCTCTC TCTCTCTCTG
25
201 CTCTCTCATC TCTCTCTCTC TNAAACNAA

TCO17(C)

1 ACAGTAGTTA GGAGTTTCTT TACTTACAAA ATCACTGGAA ATGATTAAAT
30
51 TGCTTTTCCC CCTCCCCAGA GGTGCATTTT TCTTATTTCC ATATAGTAAA
101 GTTGAGCTTT TACAGTGCAT AATGTGACAT TTGGAATGCT TATCAACTGC
151 ATGTAAACAT TAATAACCT

TCO18(C)

35
1 GTAAATGGTA TTANNNGCTG AAGAAAAAAA ATTTTCAAG ACCTCTGTTT
51 TTTAACTGAA CTTTATCATT GGCATTGTGG GCTTGAAGT TGCTGGGATA
101 AATTAATATA ATTAAATAAA AGACTGAATT TAATTGCAA AAAAAAAAAA
40
151 AACAAAGT GTGGTGAT

TCU2/1(1)

1 AAGAAATTAT CCAGTTATTT ACAAGGCCAC TGATATTTTA AACGTCCAAA
45
51 AGTTTGTTTA AATGGGCTGT TACCGCTGAG AATGATGAGG ATGAGAATGA
101 TGGTTGAAGG TTACATTTTA GGAATGAAG AACTTAGAA AATTAATATA
151 AAGACAGTGA TAAATACAAA GAAGATT

TCU2/2(1)

50
1 CGGGTTAATA TTATCCTCTA GTATAAGTGA ATTACTAGTT TCTCTTATT
51 TAGACAAACA CACACACACC AGATAATATA AACTTAATAA ATTATCTGTT
101 AATGTAGATT TTATTTAAAA AACTATATTT GAACATTGGT CTTTCTTGGA
151 C

55

TCU9/1(2)

5
 1 ACATAACAGC TTTTATACAA TGATAAGGAC ATATCATTG TTTACAAAGA
 51 AAGTCTAAAA TTTCAGAAGC ATTCAAAGAG CTAACACAGT AAAGGTCATG
 101 CAAGTTCTAG AATAGTGAAT CATGACAGAA CTCATTCATT TTATCCTTTA
 151 TCTCC

TCU9/2(2)

10
 1 AAGTATGGGT AGCTAAATTT GCATTAAATT AAAAGTACAT ATAATGCAAC
 51 ACCACTCTAC ATCTGTATAC CTACGAATGT ATGTGTACTA CACACCCTTA
 101 AAATGTTTT CAAAGTCTTA ATATATTAGA ACATGTTTC ATTTTTTCAT
 15 GGGATGTAA TACTATTCTA TGATTAAGAA AATACTAG

TCU10(2)

20
 1 AATACAGTTA TTCTAGCTTT TCATATTCAA TTTGAATGAT CAGAAAAGTA
 51 TATTAGTCAC ACAGAATTAA ATATTTTAGA TAGTAAGAAT C

TCU14(1)

25
 1 ATCCTTAGTA AGTGGATTTT GGGGAAAAAA GCACCTGGGC TTCTGGTTCT
 51 TTTTGATAAT ATATAAATT ATTCATTATG AGGTTGCAGT TGTTGCAGAA

TCU14(2)

30
 1 GAAGTGAAAG TCAGCCCTTT AGCTATTATT TATTGCTTTA TTAGAGCAGA
 51 GGGAAAGTGAC ACTCATTGCC TTCACAGAGC TCTGCAGAAA TATATGCACA
 101 GAGTGGTCAA TGCCAACATC TGAGTAAGTC TTCCCAA

TGO20(2)

35
 1 CAGAACATTA GGATTTATTC CTGATTAGT TCAAATGATT TCAACAGCTG
 51 AATTCTTTGA GATGTGTAAG GCAGGTTGGT CCTTTGGATG GACTGTAGAC
 101 TGAAACTTCC TATAACTGTA GTGATATGTA CACAGCTACA TAGCAAAGTG
 151 CTTTATTATG AAAATGAAGA A

TGO20(1)

40
 1 CAGTGTGAGA GTCTCATTTC TATGCACAGT GTTCTCAGG AGGATGGAGC
 51 TAGTTAGCTG TCTGTTGTCT GTAGCCAGC TTGATAATGG AACTATACAG
 101 CGAAGAGACA ATCTCTGGCA AGTTTTTGTA GAA

TGU5(C)

45
 1 TTAGAGTAAA ATTCCAAATA AATGCTTGC TCCAAAATTA CACTAACCAG
 51 GCTGGGTCTC TATCATACAT CTTCAATACC CTCAAACCTA GATTGTAAG
 50 101 TGAAAAAGT GATTAGCNNT TCCATTGTGTT CATTCTGTCA CTCACATTCT
 151 TAGGCATTTT AAGGATGAGC AACCTTTGTT TCAGAAAGGG TAAGTAATTA
 201 GCCCCCTGGA GGTACATAG TTATAATTTA GTCTTCAGAA TCCGTTGCAA
 251 GGGNNNGTT ACTATTTTAA AGATAATTAG AACCCACCTT GTAGCAATAA
 301 AAGTTTTCTT GTCTTTG

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TGU8(2)

1 GCGAAAGACT AATCGAACCA TCTAGTAGCT GGTTCCTCC GAAGTTTCCC
51 TCAGGATA

TGU9/1(2)

1 TTAATGTTA AATACTACTT TTTTTCAG CTTGCCCTAG ATACCAACTG
51 TTTATCTAAC ACACAATTCC AGTGTGCGA AGCCTCATGC CAATTGAAG
101 GGAACAGCCA AAACCTATGC ATTCATATAA AAAGAGTCTC TAGGCTCTTA
151 TATCTACATT ATAATTTT

TGU9/2(2)

1 GGAATAACAT TTTTATGA GGAACCCCTT TAAATGGAT GCACACAGTG
51 GCATTTTCTC CTAGGCTCAA AGCTGAGTAC ACTCCCGTAA TTTTAATAAT
101 ATTTTAGGCA AGTCCTATGA CAATTATACC AACAAGTTTC TTCAACCCCA
151 CCACCACCCC ACCATCTCTA TGC

TGU12(C)

1 GGAGGAAGCT TTATTGGGA AGAGTGGGT TCNNTCGGCC CTGATCAGCT
51 CTAGCCTGCC CACCCCATCT CAGCCAGGCG GCTTTACTTC TTCCTGAGCT
101 TCAGGTCTTT CTTCTCCTG ATTCCCTGG CCAGCTCCCC AATCAATCTC
151 CAGTACTCAT TGAACCTGAG CTCCGAGNCC TGATTCACAT CCAAGCTCTT
201 CATCTTCT

TGU13/1(C)

1 GGATGTGGTA GTTGATCTTT AATGCCCATT CTAGGTCGGA AAAATCCATG
51 ATCCTAACTT TTAAGAGAAG GTTGTAACCT CTACTTAGGA CTTTTTTTG
101 TAAGAGGAAT AATGTAGCCT CACCCCTATC TTTCTGGAAA TGTTAAACC
151 ACTGAAATAT GGAGATCAAA TCCAGCTTAC AACTGGTAA CTCAAATACT
201 ATTTTTTTTT TAACTATCT TTTCTAACT AATCACCCTT CTTGTACATA
251 GAACCTTCTA TCTCAGTGCC AATTCTTAGA GGTGATGCA AACAGCTCTC
301 CAGAGGCCT GTGCTATTGT TC

TGU13/2(2)

1 GGGGTGTACA TTTTATTGGA AACCTTAAAT ACTGTTGAGA AAGAATATAT
51 CTTCAATCAA GGTCTGCGG AGCCTACACA GAAAAATGAA GCTTTTGGG
101 TTAGGGGCAA GGATATATAC ACTACAGAGG ACAAAGA

TTO16/2(C)

1 ACATTCATTA AAGATGAACT TTCAGCATCT TCACTTGAAG ATCCATCAGA
51 TGATTCTGAG AGGCAGGTCT CCCCCAAAAA TCCACCGCAT GTATTCTTTC
101 GTTFAAATC TGAAGCCTC TTTCTTTTCA GGCTTGATGA CTCTTCTAAG
151 GTATTGTGA TGCTCTCTT CTGGGTTTTT CGTTTGCCT TATCAAGTAG
201 CTNAAATTCA AACACCATGG CAANAGAAAC TGCTTCTAT

TTO20/1(C)

5
1 CCACCAGCCT ACTGATCAGC TGGGATGCTC CTGCTGTCAC AGTGAGATAT
51 TACAGGATCA CTTACGGAGA AACAGGAGGA AATAGCCCTG TCCAGGAGTT
101 CACTGTGCCT GGGAGCAAGT CTACAGCTAC CATCAGCGGC CTTAAACCTG
151 GAGTTGATTA TACCATCACT GTGTATGCTG TCACTGGCCG TGGAGACAGC
201 CCCGCAAGCA GCAAGCCAAT TTCCATTAAAT TACCGAACAG AAATTGACAA
10
251 ACCATCCCAG ATGCAAGTGA CCGATGTTCA AGACAACTGT TTTAATAAAA
301 GATTTACATT CCAC

TTO20/2(2)

15
1 TTGGTACCAC AGTCACAGAA CTGGGGGTCA TTTTCTAGAT GAAACAAACG
51 GAACAAGTTC TCTTCCAACA AAGAAATGTA CTGTAGAAAT TAATTTCTCTC
101 CATGAATTTT ATATATTGTG TACAAATATA AGGTATGTAT CTGAATACAA
151 AG
20

TTU2/1(2)

1 CTAGAACTTC CAAAGGCTGC TTGTCATAGA AGCCATTGCA TCTATAAAGC
51 AACGGCTCCT GTTAAATGGT ATCTCCTTTC TGAGGCTCCT ACTAAAAGTC
25
101 ATTTGTTACC TAAACCTTAT GTGCCTTAAC AGGCCAATGC TTCTCG

TTU 2/2(C)

1 AACCAGTATT TCAAACTAT TATCTGGATT CAAGATTAGT GTGTAAAGAT
30
51 TGTTTTCTTA TCAGTAAAT AGGTCTTCAG ATCTGCATCT GGCCTCTTAG
101 CATGTTTTTC TTCATAGATA CCCGTTTTGG GGTTTTTGCG TCGGAAGATG
151 AAGTGCAGTT TATAGTCCTC TCCACATTTA TCTG

TTU3(1)

35
1 GGGTAGAAAG CTGAATAATT TATGAAGCAG AGGGGTCAGG GTTGATTCCG
51 GAGGACCTAT TGGTGCGGGG GCTTTGTATG ATTATGGGCG TTGATTAGTA
101 GTAGTTACTG GTTGAACATT GTTTGTTGGT GTATATATTG TAATTGAGAT
40
151 TGCTCGGGGG AATAGGTTAT GTGATTAGGA GTAGGGTTAG GATGAGTGGG
201 AAG

TTU 5/1(2)

45
1 GACAAAAAAA AAAAAACAGG TTTTAAAGCT AGAAATGAAA AGCTACTTAA
51 GTATCTTAAA GGATAAGTTA CTTTATTATA CACTAGAAAC ATACACAATA
101 GCTGAAAAC TAAAAATCT CACACTGCTG AATGTCTCTG CTGGCTG

TTU5/2(2)

50
1 GCATCCATTG TACATTGTTT GGTGTTGAGG TACCATGAGG CCTGTAAATA
51 CTATCTTATA ATTTATTATT TCAACCTGAT AAAACTTAAC ACTATTGCA
101 TAAACAAACA AACGAAAA
55

TTU9/1(1)

5 1 TAAAATACTG GTTCTTTTAT TCTGCAATAT TTTTAAAAAT CACATTTTCA
 51 GCCAGGCGCA GTTTCACACA CCTGTAATCC GGCACCTTGG GAGGCTGAGA
 101 TGGGTGGATC ACAAGGTAGG AGATCAAACA TCCTGGCCAA CATGGTGAAC
 151 CTGTTTACT

TTU9/2(2)

10 1 CAAGTATGGG TAGCTAAATT TGCATTAAAT TAAAAGTACA TATAATGCAA
 51 CACCACCTCTA CATCTGTATA CCTACGAATG TATGTGTACT ACACACCCTT
 15 101 AAATGTTTCA AAGCTTAATA TATTAGAACA TGTTTTCATT TTCAGGGAG

TTU13(2)

 1 GGAAATACAC TAGCATGTGA GCACTGTATA TAAAGCTTGA GGTTAGGAGG
 20 51 TAAAATGAAA GAAATCATTT TTAACCTCTA AGATGT

TTU13(1)

 1 TGAATTAAAT GGACTCGTTG AAAGGACAAG GAGATCGGTA ATATCTCTCT
 25 51 AAAGAACTTA TATACTAAAA TCTGTAATTG CCTGTACCAA AAGTTTGTAGT
 101 CTTCTTTT

or an analog thereof.

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11. Vector containing a DNA according to claim 10.

12. Host cell containing a vector according to claim 11.

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13. Method for isolating a gene inducible by treating chondrocytes with IL-1 β containing the steps:

- (a) hybridizing a DNA according to claim 10 under stringent conditions against DNA or RNA containing said gene; and
- (b) isolating said gene.

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14. A method according to claim 13 wherein said DNA or RNA has been isolated from chondrocytes, particularly human chondrocytes, that were treated with IL-1 β .

15. Process for expressing a gene isolated according to claims 13 or 14 containing the steps:

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- (a) cloning said gene into a suitable expression vector; and
- (b) expressing said gene in a suitable host cell.

16. Method for producing a protein containing the steps:

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- (a) culturing a suitable host cell containing a vector which contains a DNA according to claim 10 or a gene produced by a method according to claim 13 or 14; and
- (b) isolating the expressed protein.

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17. Diagnostic aid containing a DNA according to claim 10 or parts thereof or a gene isolated according to claim 13 or 14 or parts thereof.

18. Use of a DNA according to claim 10 or parts thereof or a gene isolated according to claim 13 or 14 or parts thereof for the diagnosis, prophylaxis or therapy of IL-1 β mediated diseases of connective tissues, in particular osteoarthritis or rheumatoid arthritis.

5 19. Use of a gene isolated according to claim 13 to 14 for the production of a pharmaceutical.

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